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Metal measurement in aquatic environments by passive sampling methods: lessons learning from an *in situ* intercomparison exercise

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A. Dabrin^{1*}, J-P. Ghestem², E. Uher³, J-L. Gonzalez⁴, I.J Allan⁵, M. Schintu⁶, M.J. Montero, N.⁷, J. Balaam⁸, E. Peinerud⁹, C. Miège¹, M. Coquery¹.

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 ^{1*} Irstea, UR MALY, Freshwater Systems, Ecology and Pollution research unit, 5 rue de la Doua, CS
 70077, F-69626, Villeurbanne, France
- 9 ² BRGM, 3 Avenue C. Guillemin, BP 36009, Orléans Cedex 2, France
- ³ Irstea, UR HBAN, F-92163, Antony, France
- ⁴ Ifremer, Département Biogéochimie et Ecotoxicologie, B.P. 330 Zone Portuaire de Brégaillon, 83507 La Seyne/mer cedex, France
- ⁵ Norwegian Institute for Water Research (NIVA), Gaustalleen 21, O349 Oslo, Norway
- ⁶Università degli Studi di Cagliari, Cagliari, Italy
- ⁷ AZTI, Centro Tecnológico de Investigacion Marina y Alimentaria, Marine Research Division,
 Vizcaya, Spain
- ⁸ Cefas, Centre for the Environment, Fisheries and Aquaculture Science, Lowestoft Laboratory,
 Lowestoft, United Kingdom
- ⁹ ALS Scandinavia, Box 511, SE-183 25 Taby, Sweden
- 20
- *Corresponding author :
- 22 aymeric.dabrin@irstea.fr
- 23 tel: +33 (0)4 72 20 10 53
- 24 Fax: +33 (0)4 78 47 78 75
- 25
- 26 Abstract
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Passive sampling devices (PS) are widely used for pollutant monitoring in water, but 28 29 estimation of measurement uncertainties by PS has seldom been undertaken. The aim of this work was to identify key parameters governing PS measurements of metals and their 30 31 dispersion. We report the results of an *in situ* intercomparison exercise on diffusive gradient 32 in thin films (DGT) in surface waters. Interlaboratory uncertainties of time-weighted average 33 (TWA) concentrations were satisfactory (from 28% to 112%) given the number of 34 participating laboratories (10) and ultra-trace metal concentrations involved. Data dispersion of TWA concentrations was mainly explained by uncertainties generated during DGT 35 36 handling and analytical procedure steps. We highlight that DGT handling is critical for metals 37 such as Cd, Cr and Zn, implying that DGT assembly/dismantling should be performed in very clean conditions. Using a unique dataset, we demonstrated that DGT markedly lowered the 38 39 LOQ in comparison to spot sampling and stressed the need for accurate data calculation.

⁴¹ Key words: DGT, uncertainty, blank, continental waters, coastal waters

42 Capsule: Metal concentrations obtained by DGT during an *in-situ* intercomparison exercise
43 showed that data dispersion was mainly explained by DGT handling and analytical procedure
44 steps.

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46 Highlights:

47 Uncertainties on DGT results were assessed during an in-situ intercomparison exercise48

49 DGT handling and analytical procedure steps are responsible for the dispersion of data

51 Data dispersion was similar between raw data and the TWA concentrations

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54 Introduction

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Reliable chemical monitoring of surface waters has become a crucial issue, especially in the context of the Water Framework Directive (WFD). In particular, there is an emerging need to find cheap and easy alternatives to spot sampling taking into account temporal variability (e.g. flood events, seasons). Several tools have been developed over the years to address this challenge, including bioassays/biosensors, biomonitors, chemical sensor probes and passive samplers (PS).

62 The last 20 years have seen a surge in the development of PS to enable measurements 63 of a wide range of contaminants in surface waters (e.g. Vrana et al, 2014). PS tools carry 64 many advantages, as they (i) provide an integrated measure of the contamination over the 65 period of deployment, (ii) preconcentrate contaminants, which enables a lower limit of 66 quantification (LOO) than classical analysis in spot samples and (iii) simplify the matrix of 67 the sample. Allan et al. (2006) showed that time-weighted average (TWA) concentrations 68 estimated by PS could be used to monitor long-term trends, to screen contaminants at very 69 low concentrations, and to identify sources of pollution. For metals, diffusive gradients in thin films (DGT) and Chemcatcher[®] are the main PS devices used for monitoring surface waters 70 71 (Allan et al., 2008). The DGT device consists in a plastic piston loaded with a diffusive gel 72 layer backed by an ion-exchange resin gel (Chelex-100). The diffusive gel of the DGT allows 73 to measure metals in ionic form and weakly bound to small inorganic and organic complexes, 74 which represent a fraction of the total metal dissolved concentration (<0.45 µm), commonly 75 called the "labile" fraction. Metal accumulation in the resin is governed by the concentration 76 gradient established in the diffusive gel between the media and the resin. The TWA

concentration of the metal present in water over the period of exposure may be estimated asfollows:

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$$TWAC = \frac{m \cdot \Delta g}{D \cdot t \cdot A} \tag{1}$$

80 where m is the mass of metal accumulated in the receiving phase of the DGT (ng), Δg is the 81 thickness of the diffusive gel/filter (cm), D is the diffusion coefficient of the ionic metal in the diffusive gel (cm².s⁻¹), t is the period of deployment (s), and A is the surface area (cm²) of 82 DGT. For this calculation, only *m* has to be determined by the analytical step, as Δg , *D*, *t* and 83 A are generally known (DGT supplier and field data). The Chemcatcher^(R)</sup> is constituted by a 84 85 standardized body combining a cellulose acetate membrane with a 47 mm Empore[™] 86 chelating disk for metals measurement. The uptake rates needed to calculate TWA 87 concentrations are less well known for Chemcatcher than the diffusive coefficient determined 88 for DGT (DGT Lancaster), and require element-specific uptake rate estimations in field 89 conditions (Neziri et al., 2011).

90 The DGT has a rapid response to fluctuating concentrations of Cd, Cu, Ni, Pb and Zn 91 during the period of deployment (Allan et al., 2007). For Cu, Zn and Pb, TWA concentrations 92 were in agreement with the filtered or ultrafiltered fractions obtained in spot samples and with 93 predicted inorganic and inorganic-fulvic acid associated metal fraction according to the visual 94 MINTEQ programme (Allan et al., 2007). Thus, DGT has been widely used as a speciation 95 tool for metals in freshwaters (Roig et al., 2011), coastal waters (Schintu et al., 2008), and 96 sediments (Dabrin et al., 2012). Several studies have focused on the applicability of DGT for 97 monitoring metals in freshwaters. Montero et al. (2012) reported that DGT gave reproducible 98 TWA concentrations for Cd, Cu, Ni and Zn in highly-fluctuating systems such as estuaries. 99 However, Buzier et al. (2014) reported that DGT monitoring of Cu, Cd, Ni and As in surface 100 waters requires stringent procedures to avoid DGT contamination and biases in the TWA 101 concentration estimation. Allan et al. (2008) also assessed Chemcatcher performances for 102 measuring Cd, Cu, Ni, Pb and Zn in surface waters. They reported that DGT and Chemcatcher 103 gave similar Cd and Zn concentrations to filtered samples, but both PS consistently 104 underestimated Cu and Ni concentrations and carried high measurement uncertainty for Pb 105 due to low sampler accumulation for this metal. Moreover, to improve validity of 106 Chemcatcher TWA concentrations, a performance reference compounds procedure was 107 needed to account for *in situ* turbulence, biofouling and temperature (Allan et al., 2008).

108 The upshot is that while both DGT and Chemcatcher are widely used for monitoring 109 metals in waters, there has been little effort to assess the uncertainty of their measurements.

110 To date, only one study has reported global results of an *in situ* intercomparison exercise on 111 several PS for priority pollutants (polycyclic aromatic hydrocarbons, pesticides and metals) in 112 surface waters (Miège et al. 2012). This first trial allowed to evaluate the representativeness 113 of PS for *in situ* monitoring and showed that despite the very low concentration levels, the 114 variety of tools and the different exposure and analytical strategies, global interlaboratory 115 dispersion was relatively low (relative standard deviation, RSD<100%). Nevertheless, this work needed further investigations of results on metals measurements by DGT, particularly to 116 117 scrutinize sources of TWA concentration uncertainties, such as analytical and data treatment 118 steps.

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120 **2. Strategy of the intercomparison exercise**

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Ten laboratories with previous expertise in PS deployment participated in this intercomparison exercise, i.e. ALS Scandinavia (SW), AZTI (SP), BRGM (FR), Cefas (UK), Irstea Paris (FR), Irstea Lyon (FR), EDF R&D/LNHE (FR), IFREMER Toulon (FR), NIVA (NO) and University of Cagliari (IT). Ten and 6 laboratories participated in the exercise for metals at Ternay (France) and Thau (France), respectively. Each laboratory was asked to provide its own samplers and apply its own methodology and strategy for PS assembly/deployment, PS analytical treatment and TWA concentration calculation.

Field campaigns were led on two contrasted environments: one continental (Rhône River, Ternay, France) and one coastal (Thau Lagoon, Hérault, France). All PS were deployed days at Ternay (17–24 June 2010) and 9 days at Thau (27 April–5 May 2010). Details of the exercise strategy are available in Miege et al. (2012).

133 Since some laboratories (3 at Ternay and 1 at Thau) exposed two types of PS for 134 metals, each laboratory code corresponds to the association of one laboratory and one tool. 135 The exposed PS were represented by DGT with open pores (OP) or restrictive pores (RP), while only one Chemcatcher[®] was deployed at the Ternay site (Table 1). For each laboratory, 136 137 PS were exposed in triplicate together with one field blank (i.e. brought to the field but not 138 exposed in waters). Results from PS blanks (DGT/Chemcatcher laboratory blank) and from 139 the acid used for resin elution (acid blank) were also provided by each participant. Each 140 laboratory was asked to analyze 8 metals, i.e. Cd, Pb and Ni (priority substances of the WFD; 141 EC, 2013), Cr, Cu and Zn (substances on the French ecological status list; MEEDDM, 2010), 142 Mn and Co; and to provide information related to analytical performances (Table 2), diffusion 143 coefficients used (Table 3), resin elution, analytical procedure and calculation steps (Table 3).

In order to evaluate the accuracy and uncertainty of the analytical step, all participants were provided with a reference solution (QC solution) that had to be analyzed in triplicate (n=3) at the same time as PS eluate. The reference values for the QC solution were obtained by isotope dilution coupled with inductively-coupled plasma mass spectrometry (ICP-MS). Since Mn and Co have only one isotope, a standard addition method was used.

149 In order to compare TWA concentrations against total dissolved metal concentrations, 150 surface water samples were collected at the start, during and at the end of PS deployment. 151 Samples were collected in polypropylene bottles, immediately kept at 4°C, then filtered with 152 polyvinylidene difluoride (PVDF) filters (0.45 µm) previously cleaned in HNO₃ 10% (v/v). 153 At the Ternay site, filtrates were acidified with ultra-pure HNO₃ (0.5% v/v). At the Thau site, 154 due to the saline matrix, samples were preconcentrated by liquid-liquid extraction prior to 155 analysis (Danielsson et al., 1982). All samples were then analyzed by ICP-MS (Thermo X7, Series^{II}). 156

A robust mean value and associated uncertainty was calculated for the QC solution
and the field-exposed PS according to ISO standard method 5725-5 (1994); Miège et al.,
(2012).

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- 161 **3. Results and discussion**
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163 3.1 The analytical step: the need for a clean room to avoid contaminations

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165 In order to identify which step(s) could affect the dispersion of TWA concentrations obtained by PS, a thorough evaluation of the analytical step was performed. Figure 1 displays 166 167 results of QC data from each laboratory with robust means and reference values of the QC 168 solution. The ratio between robust mean and reference value (expressed in percentage) varied 169 from 91% for Ni to 123% for Zn. Clearly, there was a lack of accuracy for Zn, since there was 170 no overlap between the reference value \pm uncertainty of the QC solution (1.030 \pm 0.036 µg/L) 171 and the robust mean $(1.263 \pm 0.130 \,\mu\text{g/L})$. This is not surprising, since Zn is known to be one 172 of the most critical metals to analyze at low levels due to sample contamination at low level. 173 Since the QC solution had to be directly analyzed without pretreatment, this suggests that 174 sample and calibration solutions have to be handled in very clean conditions (clean room 175 and/or under a laminar flow hood) to minimize contamination. A recent intercomparison 176 exercise on surface water analysis at low-level concentrations (Zn=1 µg/L) involving 24 177 routine laboratories in France also showed a tendency to overestimate Zn concentrations

(139%) (Ghestem et al., 2012). Hence, these results confirm that laboratories liable to run PS
measurements must first learn to master contamination during the analytical step, especially
for Zn.

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182 3.2 The choice of instrumental method for metal analyses: a crucial point to reduce183 uncertainties

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185 Zn, Cr, and Ni showed the highest analytical-step dispersion, at 24%, 22% and 16%, 186 respectively (Fig. 1). Our results are in accordance with the intercomparison exercise from 187 Ghestem et al. (2012) on water analysis, which also showed that at similar concentrations (Zn: 188 1 μ g/L, Cr: 1.5 μ g/L and Ni: 0.5 μ g/L), RSDs were 23%, 13% and 17%, respectively. Here, 189 two laboratories (#E and #K) reported concentrations that were systematically far from the 190 reference values for Zn, Cr and Ni. In some cases, concentrations were also far from the 191 reference values for Cd, Co, Mn, Pb or Cu, for which one or both laboratories reported results 192 with poor accuracy or poor analytical repeatability. Table 2 details the analytical technique 193 and LOQ reported by each laboratory. Most laboratories used ICP-MS, whereas laboratories 194 #E and #K used graphite furnace atomic absorption spectrophotometry (GF/AAS). As 195 expected, LOQ was one order of magnitude lower for ICP-MS than for GF/AAS (Table 2), 196 which could explain the poor repeatability and/or accuracy for GF/AAS results at the µg/L 197 level. Therefore, laboratories need to analyze PS extracts with analytical instrumentation 198 combining high precision and sensitivity to ensure valid data for low-level metal 199 concentrations in environmental waters.

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201 3.3 Blank control

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203 Figure 2 reports the results for acid blanks, DGT laboratory blanks and Ternay-site 204 DGT field blanks as the mass of analyte in the eluates. For Mn, the mass in field blanks (mean 205 of 3.6 ng) was about 1% of the mass in Ternay-exposed DGT (mean: 306 ng) after the 7-day 206 deployment. For Co, Ni, Cu and Pb, the mean contribution of mass of metal in DGT field 207 blanks to the mass of metal in exposed DGT was 11%, 12%, 12% and 21%, respectively 208 (excluding laboratory #J for Co and Pb, which reported higher values). For Zn, Cd and Cr, 209 DGT field blank levels were high compared to the mass accumulated in DGTs exposed for 7 210 days in the river, representing mean contribution of 83%, 72% and 65%, respectively. In most 211 cases, the mass of metal in DGT field blanks could be explained by a contamination already

present in DGT laboratory blanks. Since the mass of metal was negligible in acid blanks (Fig. 212 213 2), this suggests that observed contaminations were mainly due to contamination of the resin 214 itself (during its preparation) and/or to the pre-elution resin handling step. These results 215 clearly show that the determination of TWA concentrations for trace metals such as Zn, Cd or 216 Cr is a delicate issue. For Zn, high blank levels in DGT were also reported by Munksgaard et 217 al. (2003). However, Sigg et al. (2006) also reported blank values in commercially available 218 DGT for Zn which were lower (32±55 ng) than those obtained during this intercomparison 219 exercise (137±242 ng). It is possible to subtract DGT laboratory blank from results obtained for exposed DGT, but only if the results for DGT laboratory blanks are sufficiently 220 221 reproducible. To overcome this problem, a deployment over several weeks could enable to 222 trap a significantly higher mass of Zn, Cd and Cr than the mass initially present in DGT 223 laboratory blanks. Alternatively, Buzier et al. (2014) proposed to determine a LOQ-DGT 224 value representing the lowest mass of metal that could be quantified without a significant 225 blank contribution. This LOQ-DGT value was determined as the mean of DGT field blanks 226 (n=44) and 10-fold the standard deviation (mean \pm 10×SD). In our case study, this approach 227 could have been an interesting alternative but it would have required each laboratory to 228 determine its own LOQ-DGT by analyzing several DGT laboratory blanks.

Clearly, there is a need to improve the DGT measurement protocol in terms of blanks and risk of contamination. Improvement hinges on clean conditions (e.g. cleanroom, laminar flow hood), precise estimation of global contamination, and a methodology for factoring these blanks into the final result. In our opinion, the estimation of LOQ-DGT appears to be the best way forward on this issue.

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- 235 3.4 Effect of LOQ decrease
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237 One asset offered by PS is the possibility to decrease LOQ by preconcentrating the 238 metal on the receiving phase, which this intercomparison exercise has demonstrated as a 239 powerful advantage of PS devices. For Cd, for example, analytical LOQ averaged 32 ng.L⁻¹ in 240 spot samples (Table 2); but, when DGT was applied with the average elution volume (4.05 mL), average dilution factor (6.4), average diffusion coefficient of Cd at 20°C (5.3. 10⁻⁶ 241 $cm^2.s^{-1}$), average diffusive gel thickness (0.94 mm) and a period of deployment of 7 days 242 (604 800 s), the LOQ for Cd was reduced to 0.80 ng/L. This means that DGT could decrease 243 244 LOQ by a factor ~40. Another asset of PS is related to its application in coastal or marine 245 environments. Indeed, the PS devices investigated allowed to directly measure metals at ultratrace levels in coastal waters and, contrary to spot water samples direct analysis, which requires an extraction step for eliminating the saline matrix that potentially induces analytical interferences (i.e. Danielsson et al., 1982).

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250 3.5 Exploration of usual practices

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252 3.5.1 The PS extract dilution factor before analysis

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254 Table 1 shows the volume of elution and the dilution factor applied by each laboratory 255 to prepare the PS extract. These parameters ranged from 1 to 10 mL (elution volume) and 256 from 1 to 10 (dilution factor). For example, if each laboratory was handed the same PS with 257 an amount of 1 ng of metal for analysis, the actual combination of volumes of elution and 258 dilutions factor used by laboratories would have given a metal concentration in the eluate 259 ranging from 0.020 to 1 µg/L, which underlines the huge variation in laboratory practices. 260 These different practices mainly reflect laboratory habits and are also justified according to 261 their analytical LOQ, and to the percentage of nitric acid concentration required in the final 262 eluent solution. Thus, applying a small volume of eluent and minimizing the dilution factor 263 before analysis could drastically concentrate metals before analysis. The choice of the these 264 two steps also highly depends on the compromise which was chosen by each laboratory: (i) 265 obtain a more diluted sample in a large volume, which leaves the possibility to perform 266 several replicates of analysis or (ii) obtain a more concentrated sample but in a small volume; 267 thus, leaving no second chance to perform replicate analysis. These are the two crucial steps 268 in DGT extract preparation for which good practice could radically reduce problems linked to 269 contamination and decrease analytical uncertainty. These different analytical protocol-related 270 practices can lead to very different (up to a factor of 50) measured final concentrations and to 271 variation in the defined LOQ for PS.

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273 3.5.2 Correction of the PS extraction yield

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In order to evaluate the mass of metal on the resin, most laboratories applied a correction factor called "elution factor" to take into account partial extraction from the resin. As presented in Table 1, most laboratoriess applied an elution factor of 0.8, as provided by the DGT manufacturer. However, two laboratories (#C and #D) did not correct DGT data by an elution factor, and one laboratory (#K) did not correct DGT data for Cd, yet applied an elution 280 factor of 0.7 for Pb. In fact, the two laboratories that used an elution factor of 1 were those 281 that also used the highest volume of elution (10 mL). Unfortunately, this elution factor of 0.8 282 provided by the DGT supplier was mainly applied by laboratories that used the most disparate 283 conditions. Given the broad differences in elution practice, this "common" empirical 284 extraction yield emerges as a key factor of dispersion in DGT measurement (RSD of 20% 285 with or without correction, as observed in this study); which suggests the need for either intra-286 laboratory estimation of extraction yield or the development of an elution protocol leading to 287 "total extraction" of metals from the resin.

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3.5.3 Influence of PS model constants: diffusion coefficient, gel thickness, DGT surface area

291 The diffusion coefficients (D) applied by each laboratory to calculate TWA 292 concentrations (Eq.1) from DGT results are reported in Table 3. Even though the trial 293 organizer provided the mean temperature of the exposure period at both sites (19.8°C at 294 Ternay site; 17.9°C at Thau site) to all laboratories, the diffusion coefficients used by each 295 laboratory were quite variable. For example, at Ternay site for Pb and for DGT-OP, laboratory #B applied a diffusion coefficient of 6.29 . 10⁻⁶ cm²/s whereas laboratory #D used a 296 value of $10.0 \ .10^{-6} \ cm^2/s$. In fact, for both the Ternay and Thau sites, the ratio between the 297 298 minimum and maximum value of diffusion coefficients applied for a same metal ranged from 299 1.3 for Cd to 1.6 for Pb. This large variation in diffusion coefficients is explained by the fact 300 that some laboratories (i.e. laboratory #E) had prepared their own diffusive gels and 301 characterized metal diffusion coefficients for their own gel. DGT-RP was deployed by two 302 laboratories who applied a diffusion coefficient that represents 70% of the diffusion 303 coefficients used for DGT-OP (Scally et al., 2003).

304 Concerning the other coefficients of the equation (Eq.1), get thickness (Δg) values 305 applied by each laboratory were fairly similar, ranging from 0.76 mm to 0.94 mm and varying 306 according to the different DGTs used. Also, surface area of DGT (A) was similar (3.14 cm²) 307 for all DGTs purchased from DGT Research Limited, Lancaster (Table 1). However, some 308 laboratories applied different surface areas to calculate TWA concentrations for DGT-OP and 309 DGT-RP (#L and #G: 3.80 cm²; #D 4.91 cm²). The 3.80 cm² area represents the effective 310 sampling area provided by Warnken et al. (2006) when a diffusive boundary layer (DBL) is 311 taken into account; while the value of 4.91 cm² used by one laboratory (#D) is a wrong value, 312 since its represents the total surface area of the filter. In this case, the total diffusion layer 313 comprises the filter and the gel layer (Δg) plus the DBL thickness δ (0.2 mm in well-stirred

cases). Even if the surface area applied by laboratory #D was 56% higher than by the other
laboratories, the TWA concentrations calculated were not identified as outlier (Fig. 4 and 5),
suggesting that this term of the equation had no impact on the final result of this laboratory.

317 For example, we applied different combinations of data in the model calculation from 318 raw data obtained by laboratory #F for Cu at Ternay site. By applying a thickness of 0.91 mm 319 (gel + filter), a surface area of 3.14 cm^2 and by neglecting the DBL, this laboratory obtained a 320 TWA concentration of 0.305 μ g/L. If the thickness of the filter was neglected, result should 321 have been 16% lower (0.254 μ g/L). By applying a DBL of 0.2 mm and an adapted surface 322 area of 3.8 cm², TWA concentration should have been equal to 0.307 μ g/L, which is close to 323 the initial result. In an another case, if a DBL was added (0.2 mm) to the model calculation 324 without changing the surface area, TWA concentration should have been equal to $0.372 \,\mu g/L$ 325 which is 22% higher than initial result. These simulations of various combinations of data 326 inputs lead to TWA concentrations ranging from 0.254 to 0.372 µg/L, which correspond to 327 the range of data obtained at Ternay site for laboratories (Figure 4) which did not exhibit 328 outliers' for TWA data.

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330 3.6 Dispersion of PS measurements

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332 3.6.1 Comparison between concentrations in ng/tool and TWA concentrations

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334 Figure 3 gives the RSD (%) of Ternay-site and Thau-site data expressed in ng/tool and 335 for TWA (μ g/L). Considering all the sampling and analytical steps, the dispersion of data in 336 ng/tool was very satisfactory for Mn, Ni and Cu, with a RSD lower than 18% and 35% at 337 Ternay and Thau, respectively. For Co, the dispersion of data in ng/tool at Thau site was also 338 very satisfactory (14%) but climbed to an unsatisfactory 64% at Ternay. For Mn, Cu and Co, 339 good reproducibility and accuracy was reported for the QC solution (Cf. 3.1, 3.2). Moreover, 340 we observed little contamination of DGT blanks (Figure 2) for these metals, thus resulting in 341 little dispersion of TWA data. In contrast, for Pb, the RSD in ng/tool was higher, reaching 342 31% and 48% at Ternay and Thau, respectively. Also, data dispersion was higher for Zn, Cd 343 and Cr, ranging from 40% to 88% at Ternay and from 69% to 111% at Thau. The high 344 dispersions in the mass of metals in ng/tool for Zn, Cd and Cr suggest that contamination 345 issues, as previously discussed, were the main sources of dispersion in TWA concentrations.

TWA concentration (Figure 3) followed similar data dispersion patterns as ng/tool. At
 Ternay for example, RSD for Cu was 18% in ng/tool while RSD of TWA concentration was

348 15%. At Thau, the dispersion of Cd in ng/tool (111%) was close to that of TWA concentration 349 (101%). Also, data dispersion was reduced, by 21% for Co at Ternay site and by 27% for Cr 350 at Thau site, suggesting that dispersion in ng/tool could be reduced by data used as model 351 inputs. Since data dispersion was similar between data in ng/tool and in TWA concentration, 352 this suggests that whatever the data used as model inputs by laboratories, calculation was 353 proportional, dispersion of data being mainly driven by DGT handling and analytical steps. 354 Although laboratories used different values for certain parameters like diffusion coefficient, 355 gel thickness or surface area (Table 1 and 3), the calculation step did not lead to any greater 356 dispersion of the data. This suggests that the different coefficients needed to calculate TWA 357 concentration did not add significant dispersion over that calculated for DGT accumulated mass. Thus, even if diffusive coefficient values could vary (Table 3) since they are specific to 358 359 the gel used by each participant, they allowed leading to a similar TWA concentration.

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361 3.6.2 Dispersion of TWA concentrations compared to single analyses

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363 The dispersion of the TWA concentrations obtained during this intercomparison 364 exercise was compared against that obtained in one international proficiency test on 365 freshwaters spiked with selected metals (Table 4; Brunori et al., 2007). The comparison 366 showed higher RSDs for Zn (79-99%), Pb (58-112%) and Mn (28-35) measured by DGT. For 367 Zn and Pb, these higher RSDs could be explained by biases induced by the PS itself, during 368 field deployment (e.g. contamination of PS) and by the low level of concentrations in these 369 two environments (spot sampling: from 0.67 to 1.80 μ g/L for Zn and from 0.02 to 0.37 μ g/L 370 for Pb). For Mn, the RSD was lower for the proficiency testing (11%) than for this *in situ* PS 371 exercise (28%), probably since proficiency testing was led with 44-fold higher water 372 concentrations than our TWA concentrations. For Cu, Cr and Cd, PS gave similar dispersion 373 as the RSD of the proficiency testing dataset; whereas for Ni, PS gave lower dispersion (41%) 374 compared to proficiency testing (75%). To conclude, the data dispersion on TWA 375 concentration obtained during this PS intercomparison proved very satisfactory given the 376 different laboratories, different tools, in situ deployment and ultra-trace concentrations of metals in these two environments (e.g. $Cd \sim 12 \text{ ng.L}^{-1}$). 377

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379 3.6.3 Comparison of dispersion in TWA concentrations according to laboratory and type of

- 380 PS
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382 Figures 4 and 5 plot TWA concentrations obtained by each laboratory and tool at the 383 Ternay and Thau sites, respectively. Robust means of TWA concentrations obtained for all 384 laboratories are also displayed. At Ternay, only one Chemcatcher was deployed during the 385 intercomparison exercise and results were only reported for Cd, Cu, Ni, Pb and Zn. Despite 386 satisfactory QC results, TWA concentrations obtained with Chemcatcher nevertheless 387 included some outliers (Cochran test), notably for Cd, Pb and Zn. Unfortunately, with only 388 one dataset, the lack of comparison rules out any definitive conclusions on interlaboratory 389 dispersion for Chemcatcher.

390 Theoretically, DGT-RP allows the diffusion of free ions and small organic complexes, 391 while DGT-OP also allows the diffusion of metals associated with large organic ligands. In 392 fact, Allan et al. (2007) demonstrated that combining DGT-RP with DGT-OP allows to assess 393 the metal fraction linked to large organic ligands. Nonetheless, results obtained for DGT-OP 394 and DGT-RP by the same laboratory (#F and #M) at Ternay were not significantly different 395 for any of the selected metals (Figure 4). It is difficult to reach a conclusion for the other 396 laboratories (#G and #L), since in most cases (Cr, Co, Cu, Mn, Ni and Pb) TWA 397 concentrations from DGT-RP and/or DGT-OP showed high dispersion according to Cochran 398 and/or Grubbs tests. For Cd and Zn, we found no significant differences between DGT-OP 399 and DGT-RP. These results suggest that, in this specific context, it was difficult to 400 discriminate labile metals (free ions and small inorganic complexes) from metals associated 401 with large organic ligands, even when the analyses were performed by the same laboratory.

402

403 4. Conclusion

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405 This in-situ intercomparison exercise on PS for metal analysis in surface waters provided, to 406 our knowledge, a first attempt to identify key parameters controlling TWA metal 407 concentrations estimation and to quantify uncertainties and evaluate their main sources/origin. 408 We clearly demonstrated that DGT handling and analytical procedure steps are responsible 409 for the dispersion of data. Thus, it is necessary that laboratories using such passive samplers 410 for trace metal analysis master contamination by using a clean room (Class 100) for all 411 analytical steps, from the assembly of the DGT tool, their dismantling and elution, to the 412 analysis of eluates. Moreover, laboratories must analyze PS extracts with analytical 413 instrumentation combining high precision and sensitivity to ensure valid data, allowing 414 applying large combination of elution volume and dilution before analysis. For natural marine 415 and fresh waters, the use of ICP-MS is highly recommended. Finally, we showed that

416 whatever the data inputs (i.e., diffusive coefficient, elution factor from bibliography for 417 purchased DGT, or determined in laboratory for home made DGT) which were used by 418 laboratories in model calculation, data dispersion was similar between raw data obtained in

- 419 ng/tool and the TWA concentrations.
- 420

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427

428 References429

430 NF T 90-210, Protocole d'évaluation initiale des performances d'une méthode dans un
431 laboratoire – AFNOR - Mai 2009

432

Allan, I.J., Knutsson, J., Guigues, N., Mills, G.A., Fouillac, A.M., Greenwood, R., 2008.
Chemcatcher® and DGT passive sampling devices for regulatory monitoring of trace metals
in surface water. Journal of Environmental Monitoring 10, 821-829.

436

Allan, I.J., Knutsson, J., Guigues, N., Mills, G.A., Fouillac, A.M., Greenwood, R., 2007.
Evaluation of the Chemcatcher and DGT PSs for monitoring metals with highly fluctuating
water concentrations. Journal of Environmental Monitoring 9, 672-681.

440

Allan, I.J., Vrana, B., Greenwood, R., Mills, G.A., Knutsson, J., Holmberg, A., Guigues, N.,
Fouillac, A.M., Laschi, S., 2006. Strategic monitoring for the European Water Framework
Directive. Trends in Analytical Chemistry 25, 704-715.

- 445 Batley, G.E., Florence, T.M., 1976. Determination of the chemical forms of dissolved 446 cadmium, lead and copper in seawater. Marine Chemistry 4, 347-363.
- 447

Brunori, C., Ipolyi, I., Pellegrino, C., Ricci, M., Bercaru, O., Ulberth, F, Sahuquillo, A.,
Rosenberg, E., Madrid, Y., Morabito, R., 2007. The SWIFT-WFD Proficiency Testing
campaigns in support of implementing the EU Water Framework Directive. Trends in
Analytical Chemistry 26, 993-1004.

452

Bryan, S.E., Tipping, E., Hamilton-Taylor, J., 2002. Comparison of measured and modelled
copper binding by natural organic matter in freshwaters. Comparative Biochemistry and
Physiology - C Toxicology and Pharmacology 133, 37-49.

456

Buzier, R., Charriau, A., Corona, D., Lenain, J.F., Fondanèche, P., Joussein, E., Poulier, G.,
Lissalde, S., Mazzella, N., Guibaud, G., 2014. DGT-labile As, Cd, Cu and Ni monitoring in

- 459 freshwater: Toward a framework for interpretation of in situ deployment. Environmental
- 460 Pollution 192, 52-58.

461 462 Dabrin, A., Durand, C.L., Garric, J., Geffard, O., Ferrari, B.J.D., Coquery, M., 2012. 463 Coupling geochemical and biological approaches to assess the availability of cadmium in 464 freshwater sediment. Science of the Total Environment 424, 308-315. 465 466 Danielsson, L.G., Magnusson, B., Westerlund, S., Zhang, K., 1982. Trace metal 467 determinations in estuarine waters by electrothermal atomic absorption spectrometry after 468 extraction of dithiocarbamate complexes into freon. Analytica Chimica Acta, 144, 183-188. 469 470 Davison, W., Zhang, H., 1994. In situ speciation measurements of trace components in natural 471 waters using thin-film gels. Nature 367, 546-548. 472 473 DGT Lancaster - http://www.dgtresearch.com/dgtresearch.pdf 474 475 EC, 2013. Directive 2013/39/EU of the European Parliament and of the Council of 12 August 476 2013 amending Directives 2000/60/EC and 2008/105/EC as regards priority substances in the 477 field of water policy. Official Journal of the European Union. L 226: 1-17. 478 479 Forsberg, J., Dahlqvist, R., Gelting-Nyström, J., Ingri, J., 2006. Trace metal speciation in 480 brackish water using diffusive gradients in thin films and ultrafiltration: Comparison of 481 techniques. Environmental Science and Technology 40, 3901-3905. 482 483 Ghestem, J.P., Guarini, P., Charpentier, R., Labarraque, G. 2012. Intercomparaison sur 484 l'analyse des métaux dans l'eau : essai à faibles niveaux de concentration. Rapport final. 485 BRGM/RP-61841-FR, 160p. 486 487 Kottelat, R., Vignati, D.A.L., Chanudet, V., Dominik, J., 2008. Comparison of small- and 488 large-scale ultrafiltration systems for organic carbon and metals in freshwater at low 489 concentration factor. Water, Air, and Soil Pollution 187, 343-351. 490 491 Mantoura, R.F.C., Dickson, A., Riley, J.P., 1978. The complexation of metals with humic 492 materials in natural waters. Estuarine and Coastal Marine Science 6, 387-408. 493 494 MEEDDM, 2010. Arrêté du 25 janvier 2010 relatif aux méthodes et critères d'évaluation de 495 l'état écologique, de l'état chimique et du potentiel écologique des eaux de surface pris en 496 application des articles R. 212-10, R. 212-11 et R. 212-18 du code de l'environnement. 497 498 Meylan, S., Odzak, N., Behra, R., Sigg, L., 2004. Speciation of copper and zinc in natural 499 freshwater: Comparison of voltammetric measurements, diffusive gradients in thin films 500 (DGT) and chemical equilibrium models. Analytica Chimica Acta 510, 91-100. 501 502 Miège, C., Schiavone, S., Dabrin, A., Coquery, M., Mazzella, N., Berho, C., Ghestem, J.P., 503 Togola, A., Gonzalez, C., Gonzalez, J.L., Lalere, B., Lardy-Fontan, S., Lepot, B., Munaron, 504 D., Tixier, C., 2012. An in situ intercomparison exercise on PSs for monitoring metals, 505 polycyclic aromatic hydrocarbons and pesticides in surface waters. Trends in Analytical 506 Chemistry 36, 128-143. 507

508 Montero, N., Belzunce-Segarra, M.J., Gonzalez, J.L., Larreta, J., Franco, J., 2012. Evaluation 509 of diffusive gradients in thin-films (DGTs) as a monitoring tool for the assessment of the chemical status of transitional waters within the Water Framework Directive. MarinePollution Bulletin 64, 31-39.

- 512
- 513 Munksgaard, N.C., Parry, D.L., 2003. Monitoring of labile metals in turbid coastal seawater 514 using diffusive gradients in thin-films. Journal of Environmental Monitoring 5, 145-149.
- 515

ISO, 1994. ISO 5725-5. Accuracy (trueness and precision) of measurement methods and
results—Part 5: Alternative methods for the determination of the precision of a standard
measurement method, 54p.

519

Neziri, A., Lazo, P., Brummer, J., Paschke, A., 2011. New passive sampling systems for
monitoring organic micropollutants and heavy metals in the Shkodra Lake. Journal of
Environmental Protection and Ecology 12, 2218-2224.

- Nolan, C.V., Fowler, S.W., Teyssie, J.L., 1992. Cobalt speciation and bioavailability in
 marine organisms. Marine Ecology Progress Series 88, 105-116.
- Qian, J., Xue, H.B., Sigg, L., Albrecht, A., 1998. Complexation of cobalt by natural ligands in
 freshwater. Environmental Science and Technology 32, 2043-2050.
- Roig, N., Nadal, M., Sierra, J., Ginebreda, A., Schuhmacher, M., Domingo, J.L., 2011. Novel
 approach for assessing heavy metal pollution and ecotoxicological status of rivers by means
 of passive sampling methods. Environment International. 37, 671-677.
- 533
- Scally, S., Davison, W., Zhang, H., 2003. In situ measurements of dissociation kinetics and
 labilities of metal complexes in solution using DGT. Environmental Science and Technology
 37, 1379-1384.
- 537
 538 Schintu, M., Durante, L., Maccioni, A., Meloni, P., Degetto, S., Contu, A., 2008.
 539 Measurement of environmental trace-metal levels in Mediterranean coastal areas with
 540 transplanted mussels and DGT techniques. Marine Pollution Bulletin 57, 832-837.
 - 541

Sigg, L., Black, F., Buffle, J., Davison, W., Galceran, J., Gunkel, P., Kalis, E., Kistler, D.,
Martin, M., Noël, S., Nur, Y., Odzak, N., Puy, J., Van Riemsdijk, W., Temminghoff, E.,
Tercier-Waeber, M-L., Toepperwien, S., Town, R.M., Unsworth, E., Warnken, K.W, Weng,
L., Xue, H., Zhang, H. 2006. Comparison of analytical techniques for dynamic trace metal
speciation in natural freshwaters.Environmental Science and Technology, 40, 1934-1941.

- 548 Vrana, B., Klučárová, V., Benická, E., Abou-Mrad, N., Amdany, R., Horáková, S., Draxler,
 549 A., Humer, F., Gans, O., 2014. Passive sampling: An effective method for monitoring
 550 seasonal and spatial variability of dissolved hydrophobic organic contaminants and metals in
 551 the Danube river. Environmental Pollution 184, 101-112.
- 552
- Warnken, K.W., Davison, W., Zhang, H., 2008. Interpretation of in situ speciation
 measurements of inorganic and organically complexed trace metals in freshwater by DGT.
 Environmental Science and Technology 42, 6903-6909.
- 556
- Wells, M.L., Kozelka, P.B., Bruland, K.W., 1998. The complexation of 'dissolved' Cu, Zn,
 Cd and Pb by soluble and colloidal organic matter in Narragansett Bay, RI. Marine Chemistry
 62, 203-217.

560	
561	Wen, L.S., Santschi, P., Gill, G., Paternostro, C., 1999. Estuarine trace metal distributions in
562	Galveston Bay: Importance of colloidal forms in the speciation of the dissolved phase. Marine
563	Chemistry 63, 185-212.
564	
565	
566	
567	
568	
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571	
572	
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575	
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- Table 1: Type and characteristics of passive sampler and extraction steps before analysis usedby each laboratory at the two deployment sites.
- 611 (1) DGTOP: DGT with a diffusive gel with open pores (~ 5 nm), DGT-RP: DGT with a diffusive gel with
- $\begin{array}{c} 612 \\ \hline \\ 612 \end{array} \quad \text{restrictive pores } (\sim 1 \text{ nm}) \end{array}$
- 613 nd : not determined
- 614 Laboratories were identified (by *, # and \Box) when they deployed two types of tools. 615
- 616 Table 2: Analytical technique and limit of quantification (LOQ) of each laboratory. 617 Laboratories were identified (by *, # and \Box) when they deployed two types of tools.
- 618 LOQ were determined according to NF T90-210 (AFNOR, 2009) or were equal to Mean_{blank} + 619 10*SD_{blank}.
- 619 10* 620
- 621 Table 3: Diffusion coefficient used by each laboratory to calculate time weighted average 622 concentration (TWA). Laboratories were identified (by *, # and \Box) when they deployed two 623 types of tools.
- Table 4: Comparison of relative standard deviation (RSD) for metal concentrations between
 this PS intercomparison exercise at Ternay/Thau sites and the SWIFT-WFD proficiency
 testing exercise on natural waters (Brunori et al., 2007).
- 628 x*: robust mean, SD: standard deviation of the robust mean, RSD: relative standard deviation.
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631 Figure captions632

Figure 1: Metal concentration (mean \pm SD, n = 3) measured by each laboratory, with interlaboratory robust mean (dotted line) and reference concentration (solid line) for the quality control (QC) solution.

- Figure 2: Amount of metal in acid blank, DGT lab blank, DGT field blank and DGT exposed
 in situ at Ternay site after 7 days of exposure for each laboratory.
- Figure 3: Relative standard deviations (in %) of DGT data (without QC outliers) in ng/tool
 (solid line) and for TWA concentration in µg/L (grey area), for Cd, Cr, Co, Cu, Mn, Ni, Pb
 and Zn at Ternay (A) and Thau (B) sites.
- Figure 4: TWA concentration of each laboratory, robust mean (dotted lines) and spot
 sampling mean concentration (solid line) for Cd, Cr, Co, Cu, Mn, Ni, Pb and Zn at Ternay
 site. Laboratories were marked when outliers were identified by Cochran (within laboratory
 variability) and Grubbs tests (between laboratory variability) for QC and TWA data; QC data:
 Cochran test, Grubbs test TWA data: □ Cochran test, Grubbs test.
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- Figure 5: TWA concentration of each laboratory, robust mean (dotted lines) and spot
 sampling mean concentration (solid line) for Cd, Cr, Co, Cu, Mn, Ni, Pb and Zn at Thau site.
 Laboratories were marked when outliers were identified by Cochran (within laboratory
 variability) and Grubbs test (between laboratory variability) for QC and TWA data; QC data:
 Cochran test, Grubbs test TWA data: □ Cochran test, Grubbs test.
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Lab code	PS tool	Ternay Site	Thau Site	Binding agent	Gel thickness (mm)	Surface area (cm ²)	Volume elution (ml)	Volume resin (ml)	Dilution before analysis	Elution factor	Analysis
Α	DGT-OP	•	•	Chelex-100	0.94	3.14	1.0	0.15	10	0.8	ICP/MS
В	DGT-OP	•	•	Chelex-100	0.80	3.14	1.8	0.16	10	0.8	ICP/MS
С	DGT-OP	•	•	Chelex-100	0.80	3.14	10	0	5.0	1.0	ICP/MS
D	DGT-OP	•	•	Chelex-100	0.78	4.91	10	0.20	nd	1.0	ICP/MS
Е	DGT-OP	•		Chelex-100	0.76	3.14	1.0	0.16	nd	0.8	GF/AAS
F*	DGT-OP	•	•	Chelex-100	0.78	3.14	2.5	0.15	3.5	0.8	ICP/MS
G#	DGT-OP	•		Chelex-100	0.78	3.80	1.0	0.16	3.0	0.8	ICP/MS
H□	Chemcatcher	•		Empore chelating disk	nd	nd	nd	nd	10	nd	ICP/MS
I	DGT-OP	•		Chelex-100	0.90	3.14	1.0	0.16	10	0.8	ICP/MS
J	DGT-OP	•		Chelex-100	0.76	3.14	10	0.15	nd	0.8	ICP/MS
К	DGT-OP	•	•	Chelex-100	0.76	3.14	4.5	0.15	nd	0.8 (Cd: 1, Pb: 0.7)	GF/AAS
L#	DGT-RP	•		Chelex-100	0.78	3.80	1.0	0.16	3.0	0.8	ICP/MS
M*	DGT-RP	•	•	Chelex-100	0.78	3.14	2.5	0.15	3.5	0.8	ICP/MS



Table 1

		Analytical LOQ (µg/L)											
Lab code	Analysis	Cd	Cr	Co	Cu	Mn	Ni	Pb	Zn				
А	ICP/MS	0.058	0.160	-	0.066	0.210	0.086	0.044	0.80				
В	ICP/MS	0.010	0.010	0.010	0.010	0.010	0.010	0.010	0.010				
С	ICP/MS	0.002	0.010	0.005	0.10	0.030	0.050	0.010	0.20				
D	ICP/MS	0.020	0.300	0.020	0.03	0.200	0.200	0.020	0.20				
Е	GF/AAS	0.010	0.200	-	0.20	-	0.400	0.120	-				
F*	ICP/MS	0.010	0.050	0.020	0.05	0.050	0.050	0.020	0.50				
$G^{\#}$	ICP/MS	0.010	0.200	0.005	0.05	0.500	0.010	0.060	0.50				
H^{\square}	ICP/MS	0.010	0.100	0.050	0.10	0.100	0.100	0.050	0.50				
I^{\Box}	ICP/MS	0.010	0.100	0.050	0.10	0.100	0.100	0.050	0.50				
J	ICP/MS	0.100	0.500	0.500	0.50	0.500	0.500	0.100	1.00				
K	GF/AAS	0.160	0.110	0.440	1.06	0.090	0.610	0.480	6.70				
L [#]	ICP/MS	0.010	0.200	0.005	0.05	0.500	0.010	0.060	0.50				
M*	ICP/MS	0.010	0.050	0.020	0.05	0.050	0.050	0.020	0.50				

667 Table 2

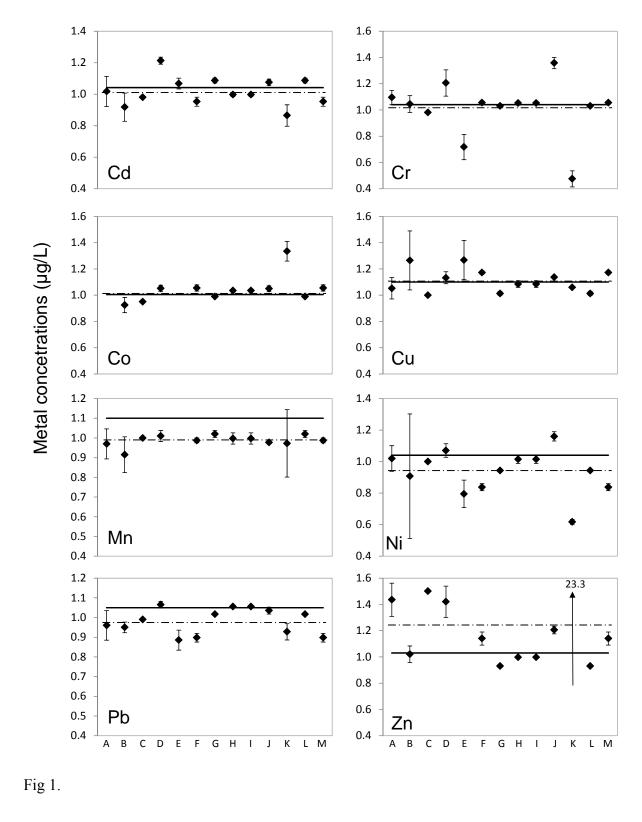
Lab code	Tool	Diffusion coefficients (10 ⁻⁶ cm ² /s)																
		С	Cd		Cr		Со		Cu		Mn		Ni		Pb		Zn	
		Ternay	Thau	Ternay	Thau	Ternay	Thau	Ternay	Thau	Ternay	Thau	Ternay	Thau	Ternay	Thau	Ternay	Thau	
А	DGT-OP	4.88	4.99	4.04	4.13	-	-	4.98	5.09	4.67	4.77	4.61	4.71	6.43	6.58	4.87	4.98	
В	DGT-OP	4.77	4.82	3.95	3.99	4.65	4.7	4.88	4.92	4.58	4.62	4.52	4.56	6.29	6.35	4.76	4.8	
С	DGT-OP	6.09	6.09	5.05	5.05	5.94	5.94	6.23	6.23	5.85	5.85	5.77	5.77	8.03	8.03	6.08	6.08	
D	DGT-OP	6.10	6.10	5.10	5.10	6.14	6.14	6.20	6.20	6.02	6.02	6.37	6.37	10.0	10.0	6.10	6.10	
Е	DGT-OP	4.86	-	4.03	-	-	-	4.97	-	-	-	4.60	-	6.40	-	-	-	
F*	DGT-OP	4.88	5.12	4.05	4.25	4.76	5.00	4.99	5.24	4.68	4.92	4.62	4.85	6.43	6.75	4.87	5.11	
$G^{\#}$	DGT-OP	6.09	-	5.05	-	5.94	-	6.23	-	5.85	-	5.77	-	8.03	-	6.08	-	
I^{\Box}	DGT-OP	4.86	-	4.03	-	4.74	-	4.97	-	4.66	-	4.60	-	6.40	-	4.85	-	
J	DGT-OP	4.86	-	4.03	-	4.74	-	4.97	-	4.66	-	4.60	-	6.40	-	4.85	-	
Κ	DGT-OP	4.88	5.12	4.045	4.25	4.76	5.00	4.99	5.24	4.68	4.92	4.62	4.85	6.43	6.75	4.87	5.11	
$L^{\#}$	DGT-RP	4.08		3.38		3.98		4.17		3.92		3.87		5.38		4.07		
M*	DGT-RP	3.41	3.60	2.83	2.98	3.32	3.49	3.49	3.67	3.27	3.44	3.23	3.39	4.49	4.72	3.40	3.58	

Table 3

Metals		In situ pa	ssive sample	r data (2010)			SWIFT-WF Exc	D proficier ercise (200	•	
	-	Гernay			Thau					
	Robust mean			Robust mean			LQ	Robust mean		
	$x^* \pm SD$	RSD	n	$x^* \pm SD$	RSD	n	Water	$x^* \pm SD$	RSD	n
	(µg/L)	(%)		(µg/L)	(%)		(µg/L)	(µg/L)	(%)	
Cd	0.005 ± 0.003	58	12	0.027 ± 0.025	92	7	0.010	0.09 ± 0.08	89	27
Cr	0.076 ± 0.070	93	11	0.036 ± 0.029	80	7	0.050	1.73 ± 1.57	91	36
Со	0.029 ± 0.011	38	9	0.060 ± 0.015	25		0.010	-	-	-
Cu	0.367 ± 0.153	42	13	0.233 ± 0.109	47	7	0.050	4.15 ± 1.66	40	42
Mn	3.47 ± 0.99	28	11	7.48 ± 2.65	35	7	0.100	154 ± 17	11	47
Ni	0.392 ± 0.139	35	13	0.261 ± 0.126	48	7	0.050	1.85 ± 1.40	75	32
Pb	0.063 ± 0.070	112	12	0.021 ± 0.012	58	6	0.010	1.20 ± 0.83	69	31
Zn	1.40 ± 1.10	79	10	3.15 ± 3.13	99	5	0.500	12.3 ± 2.8	23	39

699 700 701 Table 4

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